

IntRaFast-Q SMA NEWBORN SCREENING KIT Cat. No: 200R-40-03

PRODUCT DESCRIPTION

Spinal muscular atrophy (SMA) is characterized by degeneration of the alpha motor neurons of the spinal cord anterior horn cells, leading to progressive proximal muscle weakness and atrophy. The carrier frequency of SMA from 1/20 to 1/60. IntRaFast-Q SMA Newborn Screening Kit detects the Exon 7 and Exon 8 deletion and C/T substitution at nucleotide 840 of exon 7 in the SMN1 gene to diagnose the carrier and homozygous types by Quantitative Real Time PCR (qPCR) from dried blood spots (DBS). The kit has % 100 sensitivity and % 100 specificity for the detection of above mentioned targets in homozygous and carrier types.

PRINCIPLE OF THE SYSTEM

Test uses 5' Nuclease Assay. During the PCR reaction, the DNA polymerase cleaves the probe at the 5' end and separates the reporter dye from the quencer dye only when the probe hybridizes perfectly to the target DNA. This cleavage results in the fluorescent signal which is monitored by Real-Time PCR detection system. An increase in the fluorescent signal (Ct) is proportional to the amount of the specific PCR product. The system is based on "Intelligent Ratio (IR)" between reference and target genes quantifications. IR values may vary according to Bio-Rad CFX96 Real Time PCR devices. Therefore IntRa-Q Software should be adjusted to the Bio-Rad CFX96 Real Time PCR device during system setup (see Validation of device and Data Analysis sections).

PRODUCT SPECIFICATION

Isolated DNA should be tested with IntRaFast-Q master mix including specific primer and probes for the Exon 7 and Exon 8 deletion and C/T substitution at nucleotide 840 of exon 7 in the SMN1 gene. Please see table 1 for regions/mutations and dyes. The system provides all reagents as "ready-to-use" which has been specifically adapted for qPCR.

Table 1: Regions/mutations and Dyes

Tube	Regions / Mutations	Dyes		
IntRaFast-Q	SMN1 Exon 7	FAM		
NBS Master	SMN1 Exon 8	HEX		
Mix	Reference Gene	TEXAS RED		

SYSTEM CONTENTS

	Reagents	50 rxns	20 rxns		
•	IntRaFast-Q NBS Master Mix	1150 µl	460 µl		
•	10x Solution D	500 µl	200 µl		
•	Carrier Control DBS DNA *	30 µl	15 µl		
•	Homozygous Deletion Control DBS DNA *	30 µl	15 µl		
•	Wild-Type Control DBS DNA *	30 µl	15 µl		

^{*} Control DNAs including plasmids were adjusted to DBS extracted DNA. Amplification plots of plasmid control DNA may appear slightly different from the sample DNA.

DNA EXTRACTION

The Kit provides high sensitivity with a limit of detection (LOD) level of 0.5 $ng/\mu l$.

DBS (Dry Blood Spot)

- The system is suitable for DBS samples from newborn (heel blood etc.).
- Prepare fresh 1x Solution D by diluting the provided 10x Solution D with PCR Grade water. <u>Vortex 1-2 sn 10x Solution D before dilution.</u>
- A piece of DBS (punch-3 mm) is put into a 1.5 ml centrifuge tube.
- Vortex 1-2 sn 1x Solution D before use. Add 100 µl of 1x Solution D onto the punch. Lightly vortexed. Make sure that the punch is completely in the solution.
- Incubate for **5 minutes at room** temperature.
- 2 μI of DNA is used from the supernatant part.

VALIDATION OF DEVICE;

- Leave the master mix and control DNAs at RT to melt.
- · Mix the melted master mix gently by pipetting.
- For each control, pipette 23 μl master mix into PCR tubes/strips.
- 2 μI of different Control DNAs (carrier, homozygous deletion and wild type) and 5-10 unknown DNAs belonging to the isolation system to be studied are added to each tube to control the isolation system. Optical caps are closed. Run with the programme shown below.
- Control samples should identify expected genotypes for device validation.
 If they are not, please contact to manufacturer for validation of your device (tech@snp.com.tr).
- It is sufficient that the validation of each device should be made <u>only</u> once for each lot.
- After the device validation, continue with the standard test protocol.

STANDARD TEST PROTOCOL;

- Leave the master mix at RT to melt.
- Mix the melted master mix gently by pipetting.
- For each samples, pipette 23 μl master mix into each PCR tubes/strips.
- Add 2 μl Sample DNA into each tube and close the optical caps.
- Run with the programme shown below.

PCR PROGRAMME

96 ℃	5 Sec.	25 Oudes
60 °C	40 Sec.	35 Cycles

Select FAM, HEX and TEXAS RED as fluorescent dyes.

*The number of cycles may vary depending on the device used.

Real Time PCR time is **60 minutes for DBS** samples.



- The kit should be only used with the Bio-Rad CFX96 Instrument for detection of SMN1 Exon 7 and Exon 8 Wild-Type / Carrier and Homozygous deletion due to Intra-Q software compatibility.
- For the purpose of only detecting SMN1 Exon 7 and Exon 8 wild-type and Homozygous deletion, when SMN1 carrier analysis is not required, the kit can be used with all devices with FAM, HEX and Texas RED filters.

DATA ANALYSIS

- The threshold value should be set to 1000 for all dyes.
- The ct value for the Reference Gene (Texas RED) should be ≤ 33. Samples
 that do not comply with this value should be repeated.
- You should use IntRa-Q Software that can calculate IR values for data analysis. Please check the manual of IntRa-Q Software.
- The results should be seen like Figure 1.
- You can also check the results by amplification plots (Figures 2 to 4).

	1	2	3	4	5	6	7	8	9	10	11	12
Α	A01	A02	A03	A84	A05	A05	A07	A08	A09	A10	A11	A12
	Wild Type	Carrier	Wild Type	Wild Type	Wild Type	Wild Type	Wild Type	Wild Type				
	2.62	2.88	2.91	3.13	3.13	8.53	3.08	3.18	2.67	2.94	3.13	4.26
В	B03	B02	B01	B04	B05	B06	B07	B08	B09	B10	B11	B12
	Wild Type	Wild Type	Carrier	Wild Type	Wild Type	Wild Type	Wild Type	Wild Type				
	2.61	2.84	9.59	3.44	3.16	3,33	3.65	3.07	1.82	2.70	2.79	3.87
С	C01 Wild Type 2.98	C02 Wild Type 3.26	C03 Wild Type 3.41	C84 Wild Type 2.83	C05 Wild Type 2.70	C06 Wild Type 3.17	C07 Wild Type 3.42	C08 Homozygous Deletion -1	C09 Wild Type 3.30	C10 Wild Type 3.27	C11 Wild Type 3.83	C12 Wild Type 3.91
D	D01	D02	D03	D84	D05	D06	D07	D08	D09	D10	D11	D12
	Wild Type	Carrier	Wild Type	Wild Type	Wild Type	Wild Type	Wild Type					
	3.27	9.10	2.03	2.99	3.08	3.49	3.87	3.71	3.77	3.16	3.37	4.13
E	E01	E02	E03	E04	E05	E06	E07	E08	E09	E10	E11	E12
	Wild Type	Carrier	Wild Type	Carrier	Wild Type	Wild Type						
	2.84	3.27	3.21	2.70	3.08	3.28	3.38	10.08	3.47	9.84	3.14	3.89
F	F01	F02	F03	F04	F05	F06	F07	F08	F09	F10	F11	F12
	Wild Type	Wild Type	Wild Type	Carrier	Wild Type	Wild Type	Wild Type	Wild Type	Wild Type	Wild Type	Wild Type	Wild Type
	3.44	3.13	3.06	9.04	3.59	3.21	3.30	3.76	2.73	2.77	3.16	3.42
G	G01	G02	G03	G04	G05	G06	G07	G08	G09	G10	G11	G12
	Wild Type	Wild Type	Wild Type	Wild Type	No DNA	Homozygous	Wild Type	Wild Type	Repent	Wild Type	Wild Type	Wild Type
	2.86	3.36	1.72	3.04	-2	Deletion	3.47	1.80	53.76	2.94	3.09	3.36
н	H01	H02	H03	H84	H05	H06	H07	H08	H09	H10	H11	H12
	Homozygous	Wild Type	Wild Type	Wild Type	Wild Type	Wild Type						
	Deletion	2.85	2.99	2.47	4.42	3.51	3.09	2.73	1.21	2.68	2.93	2.91

Figure 1: Results by IntRa-Q Software

IntRa-Q Software is based on the IR method based on "reference gene quantitation/ SMN1 gene Exon 7 and Exon 8 quantitation" ratio calculation. The quantitation values of the reference and SMN1 gene are calculated with the slope curve embedded in the software, and thus the IR value is determined by the ratio of these values to each other. The software shows the SMA test as carrier, homozygous deletion and wild-type depending on these numerical IR values⁽¹⁾.

To check the amplification plots of SMN1 Exon 7 and Exon 8 wild type, carrier states and homozygous deletion, FAM (SMN1 Exon 7 - blue plot), HEX (SMN1 Exon 8 - green plot) and TEXAS RED (Reference gene - red plot) dyes should be analyzed together.

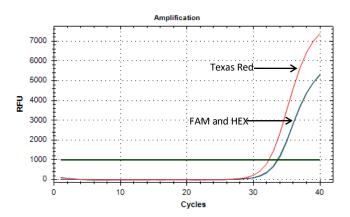


Figure 2: SMN1 Exon 7 and Exon 8 Wild-Type Sample

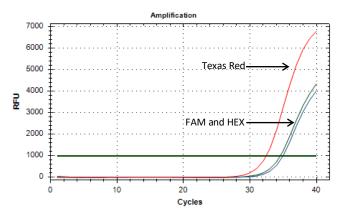


Figure 3: SMN1 Exon 7 and Exon 8 Carrier Sample

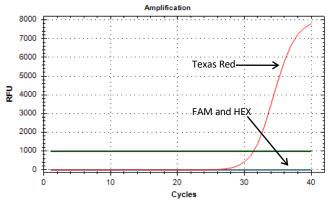


Figure 4: SMN1 Exon 7 and Exon 8 Homozygous Deletion Sample



Important points to take into account;

- The results of homozygous deletion and carrier state should be checked from the data analysis screen as amplification plots.
- Some PCR failure problems; like air bubble formation, pipetting etc. may
 cause unaccepted amplification plots as seen in Figure 5, 6, 7 and 8 it may
 not be evaluated properly by the software. Please check the amplification
 plots and re-test the sample if required.
- If the sample is SMN1 Exon 8 carrier and Exon 7 wild-type states, it should be re-tested. In case the results are same, it should be confirmed by DNA sequence analysis.
- If the sample is SMN1 Exon 7 carrier and Exon 8 wild-type states, it should be re-tested for confirmation.

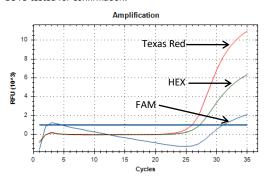


Figure 5: Unaccepted amplification plots

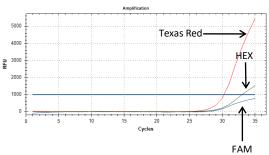


Figure 6: Unaccepted amplification plots

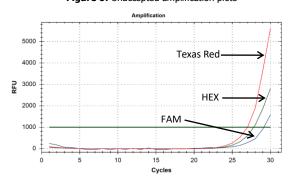


Figure 7: Unaccepted amplification plots

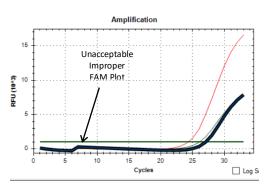


Figure 8: Unaccepted amplification plots

Inability in the IntRaFast-Q SMA Screening Analysis;

- The IntRaFast-Q SMA Newborn Screening Kit performs carrier state and homozygous deletion on Exon 7 and Exon 8 of SMN1 gene. Therefore, the kit does not detect other rare intragenic mutations (2-4%) that cause SMA disease.
- The IntRaFast-Q SMA Newborn Screening Kit detects the dosage of SMN1 gene. In a normal individual, 2 copies of the SMN1 gene are found (1 + 1). In rare cases (1-4%), there is no SMN1 gene in one chromosome but 2 SMN1 genes in the other chromosome (2 + 0). In this case, two copies of SMN1 are reported with the kit. However, the subject is the carrier state for SMA disease.

TROUBLE SHOOTING

If there is no amplification in the well,

- Absence of DNA
- Sample is containing DNA inhibitor(s)

Please contact us for your questions, tech@snp.com.tr

CAUTIONS

- All reagents should be stored at suitable conditions.
- $\bullet\hspace{0.4cm}$ Do not use the PCR master mixes forgotten at room temperature.
- Shelf-life of PCR master mix is 24 months. Please check the manufacturing date before
- Only use in vitro diagnostics and research.

STORAGE

- All reagents should be stored at 20 °C and dark.
- All reagents can be used until the expiration date, mentioned on the box label.
- Repeated thawing and freezing (>5X) should be avoided, as this may reduce the sensitivity of the assay.

REFERENCES

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