SNP Biotechnology R&D Ltd.
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Y CHROMOSOME MICRODELETION REAL-TIME PCR KIT (8 REGIONS)
Cat. No: 15R-10-08

INTRODUCTION
Microdeletions of the Y chromosome are a recently discovered cause of spermatogenetic failure resulting in male infertility. In the last decade, many investigators have described the occurrence of microdeletions in infertile patients around the world. The molecular detection of deletions has become an important diagnostic test in male infertility studies. Three Azoospermia factors (AZFa, AZFb and AZFc) have been mapped to Yq11, of which AZFc is the most frequent region involved in deletions [1,2].

INTENDED USE
Y Chromosome Microdeletion Real-Time PCR Kit (8 regions) can detect deletion of the four Azoospermia factors (AZFa, AZFb and AZFc) on Y Chromosome in whole blood samples by using qualitative Real-Time PCR method.

TARGETED USER
For professional use only. Testing should be performed by professionals trained in molecular techniques.

PRINCIPLE OF THE SYSTEM
During the PCR reaction, the DNA polymerase cleaves the probe at the 5′ end and separates the reporter dye from the quencer dye only when the probe hybridizes perfectly to the target DNA. This cleavage results in the fluorescent signal which is monitored by Real-Time PCR detection system. An increase in the fluorescent signal (C_T) is proportional to the amount of the specific PCR product [3,4].

PRODUCT SPECIFICATION
Each isolated DNA should be tested with all master mixes separately. The kit provides reagents in a **"ready-to-use"** master mix format which has been specifically adapted to 5′ nuclease PCR for SNP analysis. The test system is designed by SNP Biotechnology for use with sequence specific primers and probes.
The fluorescence of AZF regions analysis is FAM dye. Also each master mix contains an internal control labelled with HEX/JOE dye. Internal Control is Prothrombin gene – FII (OMIM: 176930).

SYSTEM CONTENTS

Reagents	10 rxns	20 rxns	50 rxns
ZFY/X (Control) Master Mix	200 µl	400 µl	1000 µl
SY14 – SRY (Control) Master Mix	200 µl	400 µl	1000 µl
SY84 - USP9Y (AZF-a)	200 µl	400 µl	1000 µl
SY86 (AZF-a)	200 µl	400 µl	1000 µl
SY127 (AZF-b)	200 µl	400 µl	1000 µl
SY134 (AZF-b)	200 µl	400 µl	1000 µl
SY254 (AZF-c/DAZ)	200 µl	400 µl	1000 µl
SY255 (AZF-c/DAZ)	200 µl	400 µl	1000 µl
Male Normal Control DNA*	100 µl	100 µl	200 µl
Female Control DNA*	100 µl	100 µl	200 µl

Table 1: Kit content

*Since to Control DNA is a synthetic plasmid, amplification plots of synthetic control DNA may appear slightly different from the sample DNA. Amplifications of control DNAs can be found in Table 4. Please gently vortex and then spin centrifuge for 1-2 seconds before use the control DNAs.

STORAGE
- All reagents should be stored at – 20 °C and dark.
- All reagents can be used until the expiration date on the box label.
- Repeated thawing and freezing (>4X) should be avoided, as this may reduce the sensitivity of the assay.

SAMPLE COLLECTION
Y Chromosome Microdeletion Multiplex Real-Time PCR Kit (8 regions) is approved for use with whole blood samples.
- Standard precautionary instructions must be followed by all healthcare professionals during the collection and transportation of whole blood samples.
- Whole blood samples should be collected in appropriate containers before delivery to the laboratory.
- Freezing and thawing of samples should be avoided.

DNA EXTRACTION
Blood samples should be collected in appropriate sterile EDTA tubes and can be stored at +4°C up to one month. For more than one month specimen should be stored at -20°C. It is advised to gently mix the tube (with EDTA) after collection of blood to avoid coagulation.
Our system optimized according to GeneAll® Exgene™ Blood SV. It is advised to elute DNA with 150 µl elution buffer for better results.

PROCEDURE
- Different test tubes should be prepared for each master mix.
- Leave the master mixes* and controls at RT to melt.
- Before starting work, mix the master mixes gently by pipetting
- For each sample, pipet **20 µl master mix** with micropipets of sterile filter tips to each optical white strips or tubes.
- Add **5 µl DNA** into each tube. Please do not pipette DNA before and after addition into well.
- Optical caps are closed, it is recommended to spin the plates/strips at low speed for a short time.
- Run with the programme shown below.

*Master mixes include HotStart Taq DNA Polymerase.

PCR PROGRAMME

95 °C	3 Min.	Holding
95 °C	15 Sec.	30 Cycles
60 °C	1 Min.	

Table 2: PCR Programme

Fluorescent dyes are FAM and HEX/JOE.

<u>This system can be used with the following devices ;</u>
- Bio-Rad CFX96, Opus 96
- ABI Prism ® 7500/7500 Fast
- Mic qPCR Cycler

For other two or more channel Real-Time PCR devices (which can read FAM and, HEX/JOE dyes), a trial run is recommended.

<u>If you use;</u>
ABI Prism® system, please choose **"none"** as passive reference and quencher.
Mic qPCR Cycler, please adjust gain settings, **"Green Auto Gain"** to **20** and **"Yellow Auto Gain"** to **10.**

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Supplied Materials
- White PCR plates/strips with optical covers*

*The PCR Plate/strip tube and caps seriously affect the amplification curve quality. Therefore, white PCR plates/strips and optical caps provided by the manufacturer should be used with the kit.

Required Materials (Not Provided)
- PCR Cabinet
- Vortex Mixer
- Desktop Microcentrifuge (For 2.0ml tubes and PCR strip tubes), plate spin for studies using PCR plates.
- Automated or spin column based DNA isolation Kit
- Disposible powder-free laboratory gloves
- Micropipettes (0.5ml-1000ml)
- Micropipette tips
- Standard laboratory equipments.

DATA ANALYSIS

After the run is completed data are analysed using the software with FAM and HEX/JOE dyes. The below results were studied with Bio-Rad CFX96. The threshold values for all dyes were set to 500, based on experiments conducted using the Bio-Rad CFX96 Real-Time PCR system, the GeneAll® Exgene™ Blood SV Isolation Kit, and white PCR strips supplied by SNP Biotechnology. Threshold values may vary depending on the PCR device, DNA isolation kit, and the type or brand of PCR strips/tubes used.
Internal control amplification plots must be seen in all wells except NTC and has been labelled with HEX/JOE dye. The C_T value of internal controls should be **$20 \leq C_T \leq 27$**. These values are optimised according to the GeneAll® Exgene™ Blood SV Isolation Kit and Bio-Rad CFX96 Real-Time PCR Device. C_T values may vary ±2/3 cycle according to the other DNA isolation systems and Real-Time PCR devices (Figure 1).

The C_T values range in Y chromosome regions should be **$20 \leq X \leq 27$** for valid amplifications (Figure 2-3). The absence of amplification is considered **"deletion positive"** for the relevant region. Please see table 3 for example evaluation.

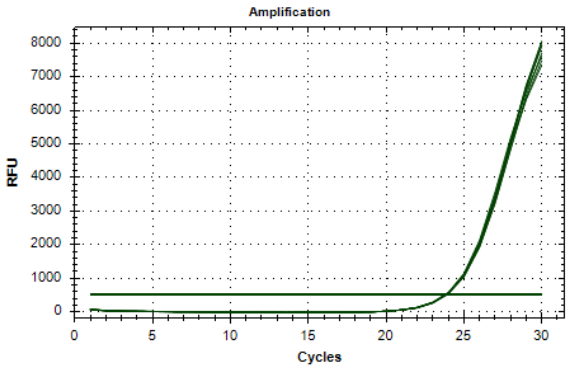

Figure 1: Internal Control plots – HEX/JOE Dye

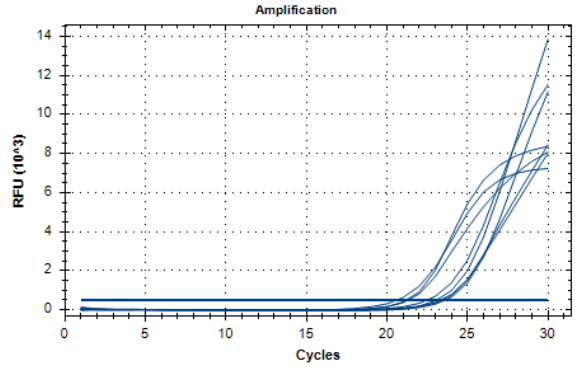

Figure 2: Normal Male Sample (FAM,Dye)

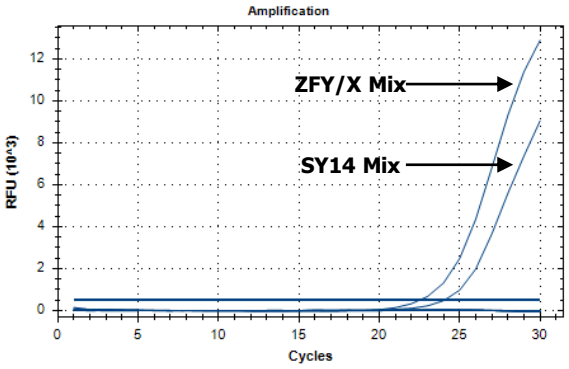

Figure 3: Male Sample for AZFa – AZFb – AZFc Deletion Positive (FAM Dye)

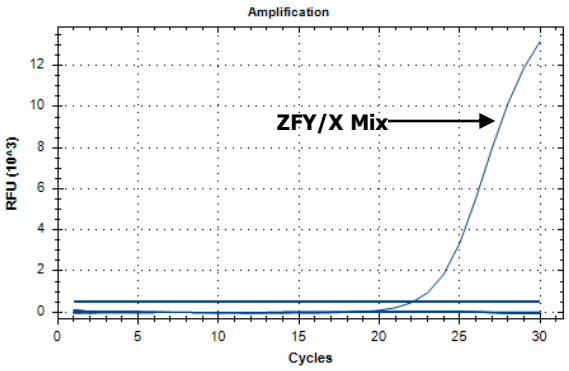

Figure 4: Female Sample (FAM Dye)

CAUTIONS
- All reagents should be stored at suitable conditions.
- Do not use the PCR master mixes forgotten at room temperature.
- Thaw PCR master mix at room temperature and slowly mix by inverting before use.
- Shelf-life of PCR master mix is 12 months. Please check the manufacturing data before use.
- Only use in vitro diagnostics.

DISPOSAL OF KIT

Dispose of it according to the legal regulations of your region

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EVALUATION OF RESULTS ACCORDING TO AMPLIFICATIONS

Dyes	ZFY/X (Control) Master Mix	SY14-SRY (Control) Master Mix	SY84 (AZFa) USP9Y Master Mix	SY86 (AZFa) Master Mix	SY127 (AZFb) Master Mix	SY134 (AZFb) Master Mix	SY254 (AZFc/DAZ) Master Mix	SY255 (AZFc/DAZ) Master Mix	Result Evaluations*
HEX/JOE (IC)	+	+	+	+	+	+	+	+	Deletion negative (Normal) Male DNA
FAM	+	+	+	+	+	+	+	+	
HEX/JOE (IC)	+	+	+	+	+	+	+	+	Female DNA
FAM	+	-	-	-	-	-	-	-	
HEX/JOE (IC)	+	+	+	+	+	+	+	+	AZFa Deletion Male DNA
FAM	+	+	-	-	+	+	+	+	
HEX/JOE (IC)	+	+	+	+	+	+	+	+	AZFb Deletion Male DNA
FAM	+	+	+	+	-	-	+	+	
HEX/JOE (IC)	+	+	+	+	+	+	+	+	AZFc Deletion Male DNA
FAM	+	+	+	+	+	+	-	-	

Table 3: Evaluation of results according to amplifications.

* Blue boxes indicate no amplification . These results are based on the most common deletions. Samples may give different amplifications from these conditions.

TROUBLESHOOTING PROBLEMS AND SOLUTIONS

Problem	Reason	Solution
Internal control does not work/ low amplification	Absence of Sample / not added into well	Repeat test
	Sample is containing PCR inhibitor(s)	
No target /internal control amplification curves in all wells	Error in temperature/time settings in PCR program	Correct any errors in the temperature/time settings in the PCR Program and repeat the test.
	Sample is containing PCR inhibitor(s)	Repeat test
Positive control result and/or C_T values are lower or higher than the value mentioned in User Manual.	Error in temperature/time settings in PCR program	Correct any errors in the temperature/time settings in the PCR Program and repeat the test.
C_T values are not valid (higher or lower) according to User Manual	Excessive or insufficient sample	Repeat the test.
Low and/or invalid amplification curves	Stability problems arising from repeated thawing and freezing (>4X)	Repeated thawing and freezing (>4X) should be avoided, as this may reduce the sensitivity of the assay.
	Sample is containing PCR inhibitor(s)	Repeat the test.
	Stability problems arising from unavailable storage conditions.	All reagents should be stored at – 20 °C and dark.
	Bubble formation or pipetting error during pipetting	After adding the master mix and sample, it is recommended to spin the plates/strips at low speed for a short time.
For further questions, please contact us **tech@snp.com.tr**		

Table 4: Troubleshooting problems and solutions

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REFERENCES

1. David S. Cram, Kun Ma, Shalender Bhasin, Jose Arias, Marintan Pandjaitan, Brendan Chu, Pam Audrins, Doug Saunders, Frank Quinn, David deKretser, and Robert McLachlan. "Y chromosome analysis of infertile men and their sons conceived through intracytoplasmic sperm injection: vertical transmission of deletions and rarity of de novo deletions". Fertility and Sterility,Vol:74, No:5, November 2000.
2. Abhijit Ray, Arunabha Tapadar, Maitreyee Kar, Rajib Kundu and Shantanu Nandy. Microdeletions in the Y chromosome in cases of male infertility in a population in West Bengal. Journal of the Anatomical Society of India. Volume 63, Issue 1, June 2014, Pages 52-56.
3. Yolanda S Lie and Christos J Petropoulos. "Advances in quantitative PCR technology: 5' nuclease assays". Current Opinion in Biotechnology Volume 9, Issue 1, February 1998, Pages 43-48.
4. Luis Ugozzoli and R. Bruce Wallace. "Allele-Specific Polymerase Chain Reaction". A Companion to Methods in Enzymology Vol. 2, No. 1, February, pp. 42-48, 1991.

SYMBOLS AND DESCRIPTIONS

REF	Catalog Number	CE	CE Mark
LOT	Lot Number	UDI	Unique Device Identifier (01)Device Identifier (17)Expiry Date (10)Lot Number
	Manufacturer	Σ	Test Quantity
	Fragile	-20 °C	Storage Temperature
	Protect from directly sunlight	IVD	In Vitro Diagnostics
	Expiry Date		

Table 5: Symbols and descriptions